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SYNTHESIS OF A GLYCOTETRAOSYL SERINE, A PARTIAL STRUCTURE OF AN OVARIAN CYST MUCIN GLYCOPROTEIN OF BLOOD GROUP A ACTIVITY¹

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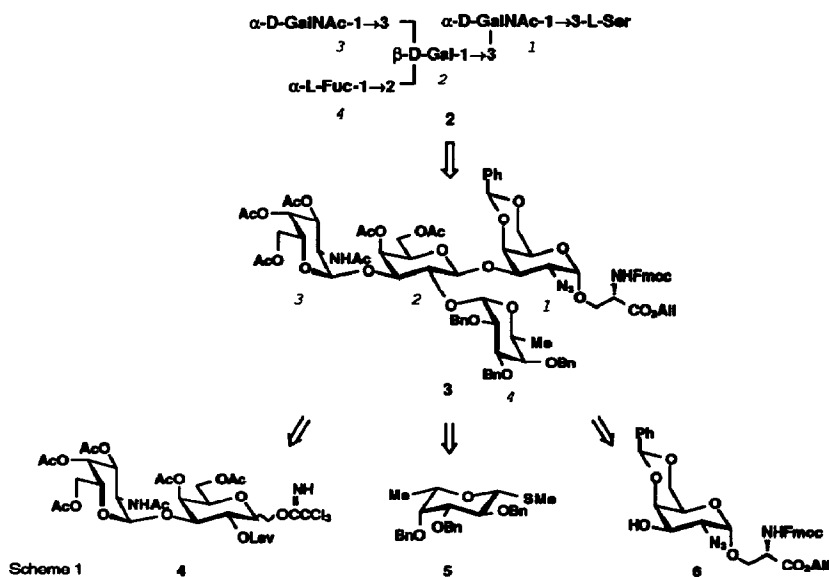
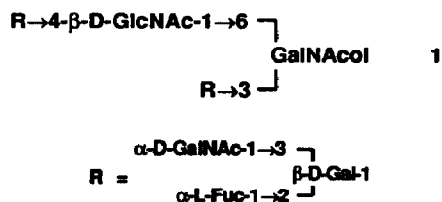
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Abstract: A glycotetraosyl serine containing blood group A determinant tetrasaccharide and its properly protected equivalent were synthesized in a regio- and stereocontrolled manner.

In 1986, Bush and his co-workers² reported the isolation of complex oligosaccharide alditols such as **1** after alkaline borohydride degradation of ovarian cyst mucin glycoproteins with blood group A activity and their characterization through a combined use of high pressure liquid chromatography and ¹H NMR spectroscopy. It is to be noted that structure **1** bears two blood group A determinant trisaccharide composite units α -D-GalNAc-(1→3)-{ α -L-Fuc-(1→2)-} β -D-Gal and belong to the core II type structure³.

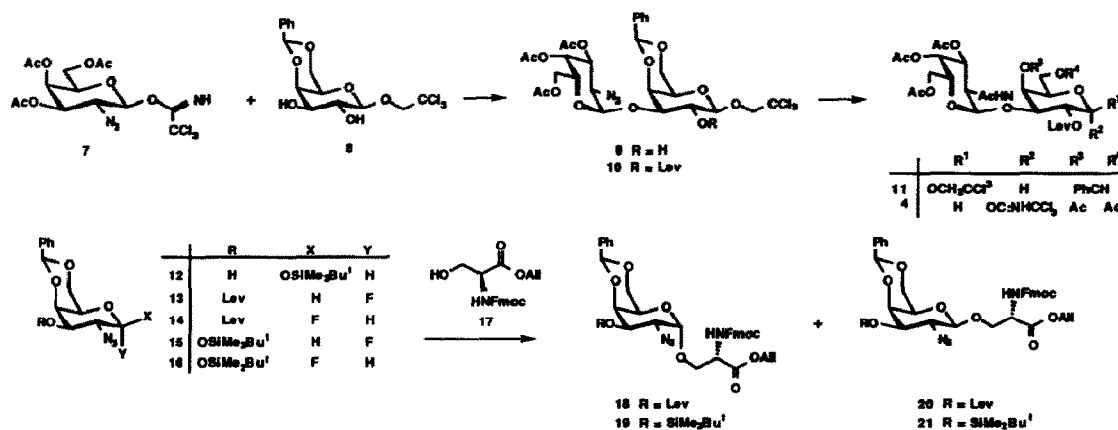
In connection with our on-going project on the synthesis of glycopeptide fragments of glycophorin, we have been interested in the development of general synthetic approaches toward O-linked glycan of glycoproteins with blood group activity. In order to study a plausible synthetic strategy directed towards highly branched structures such as **1**, a partial structure of **1** was extracted and designed as a target molecule **2** that contains L-serine as a possible linking residue of the carbohydrate periphery to the peptide backbone. Hitherto synthetic studies towards blood group A active oligosaccharides that carry no amino acid have been reported from



several groups⁴. Herein we report on the stereoselective synthesis of compounds 2.

Retrosynthetic analysis of 2, which belongs to the core I type structure, was carried out by first designing a completely protected glycotetraosyl serine 3 which should be suitable for further elaboration at O-6' of 3 in order to approach a glycosyl serine with core II type structure such as 1. Disconnection of 3 led us to design two glycosyl donors 4 and 5, and a glycosyl acceptor 6.

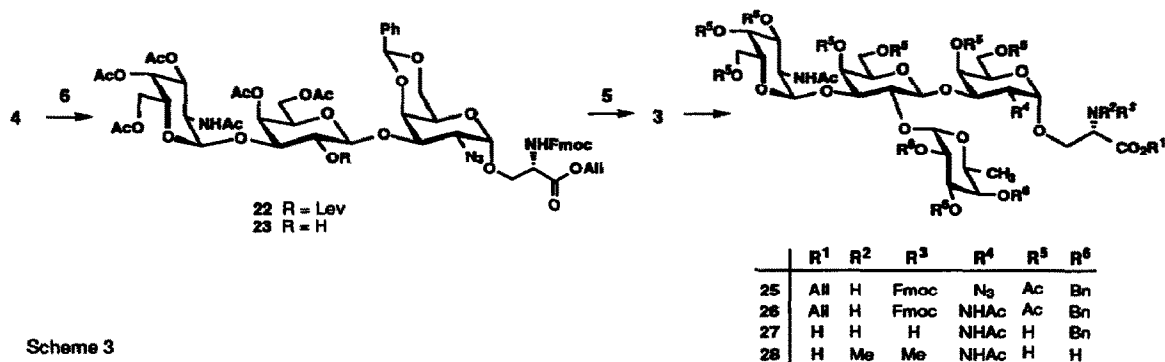
The glycosyl donor 4 was synthesized in a following way. Glycosylation of diol 8⁵ with 1.2 equivalents of β -trichloroacetimidate 7⁶ in the presence of powdered molecular sieves 4A (MS4A) and TMSOTf⁷ in CH₂Cl₂ at -20° proceeded smoothly to give a regioselectively glycosylated product 9⁸ in 77% which was treated with levulinic anhydride⁹ and DMAP in pyridine to give 86% of 10⁸. Conversion of 10 into 4⁸ was performed in 4 steps; 1) AcSH¹⁰ in CH₂Cl₂, 4 days at 20°, 2) 8:2:1 CF₃COOH-H₂O-CH₂Cl₂, overnight at 20°, 3) Ac₂O, DMAP in pyridine, 4) Zn, 2:5 AcOH-THF, 5) CCl₃CN¹¹, DBU in CH₂Cl₂, 83% overall.



Scheme 2

The glycosyl acceptor 6 was prepared as follows. A readily available 12¹² was converted into α - and β -fluorides 13⁸ and 14⁸ in 40% and 35% overall yield in three steps; 1) (Lev)₂O, DMAP in pyridine, 2) nBu₄NF, AcOH in THF¹³, 3) DAST¹⁴ in CH₂Cl₂. Glycosylation of the serine derivative 17^{15,20} with 0.4 equivalents of 13 in the presence of Cp₂Zr(ClO₄)₂¹⁶ and MS4A in CH₂Cl₂ afforded 46% of desired 18⁸ and 28% of crude β -isomer 20. A similar observation was made under the same reaction condition by use of β -fluoride 14 and 52% of α -isomer 18 was obtained along with 21% of crude β -isomer 20. Compounds 13 and 14 was then converted into another pair of glycosyl donors 15⁸ and 16⁸ in 90 and 89% yield, respectively, in two steps: 1) NH₂NH₂AcOH in 1:5 PhMe-EtOH¹⁷, 2) ^tBuMe₂SiCl, imidazole, DMAP in DMF. Now glycosylation of 17 with 0.96 equivalents of 15 according to the Suzuki procedure as described above gave an improved result and thus a 93% yield of mixture 19⁸ and 21⁸ in a ratio of 10:1 was obtained. Use of the β -fluoride 16 gave virtually the same result. Treatment of 19 with NH₄F¹⁸ in MeOH afforded the designed glycosyl acceptor 6 in 72% yield. TMSOTf catalyzed coupling of 4 with 1.3 equivalents of 6 in the presence of MS4A in CH₂Cl₂ for 12h at -20° gave 58% of 22⁸. Removal of levulinoyl group of 22 was carried out as described above to give 98% of 23⁸. Crucial glycosylation of 23 with 10 equivalents of a fucosyl donor 5 was achieved in the presence of

$n\text{Bu}_4\text{NBr}\cdot\text{CuBr}_2\cdot\text{AgOTf}^{19}$ in 5:1 $(\text{CH}_2\text{Cl})_2\text{-PhMe}$ to give 58% of the designed key intermediate 3. The β -isomer⁸ of 3 was also formed and was isolated in 14% yield.



Scheme 3

Finally, in order to confirm the structure, completely protected 3 was converted into free glycotetraosyl serine derivative 28⁸ via 26⁸ and 27⁸ in 5 steps: 1) 80% aq.AcOH, 6h at 60°, then Ac₂O, DMAP in pyridine, 87%; 2) AcSH and pyridine, 94%; 3) Pd(Ph₃P)₄, PhNHMe in THF²⁰, 74%; 4) aq. NaOH-MeOH, 95%; 5) 20% Pd(OH)₂-C, H₂, 80% aq.MeOH, 89%. Hydrogenolysis of three benzyl groups on fucosyl residues of compound 27 in step 5 did afford N,N-dimethylated serine derivative 28 instead of the expected 2, most probably due to the presence of trace amount of HCHO in the reaction medium. Since structure of 28 was firmly confirmed by ¹H-NMR and FAB-MS data, we are now in a position to extend our synthetic strategy directed further toward glycosyl serines with core II type structure.

In summary, by employing two glycosyl donors 4 and 5 as well as a glycosyl acceptor 6, a rational synthetic design to core I type glycosyl serine with blood group A activity was developed.

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References and Notes

- Part 96 in the series "Synthetic Studies on Cell-Surface Glycans". For part 95, see F. Goto and T. Ogawa, *Tetrahedron Lett.*, submitted.
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8. Physical data for novel compounds are presented below. Values of $[\alpha]_D$ and $\delta_{H,C}$ were recorded at $25 \pm 3^\circ$ for solutions in CHCl_3 and CDCl_3 , respectively, unless otherwise indicated. Signal assignment such as H-3² stands for a proton at C-3 of sugar residue 2. 3: $[\alpha]_D +51.6^\circ$ (c 0.5); δ_H 0.992 (d, J 6.6 Hz, Fuc-CH₃), 1.917, 1.935, 1.965, 1.997, 2.063, 2.096 (6s, 6Ac), 5.158 (d, J 3.7 Hz, H-1³), 5.438 (d, J 3.7 Hz, H-1⁴), 5.557 (s, PhCH), 5.645 (d, J 9.2 Hz, SerNH or NHAc); δ_C 94.5 (C-1³), 98.1 (C-1⁴), 100.6 (C-1⁷), 101.5 (PhCH). FAB-MS (M+1)⁺ 1634. β -isomer of 3: $[\alpha]_D +4.6^\circ$ (c 0.2); δ_H 1.010 (d, J 6.2 Hz, Fuc-CH₃), 1.811, 1.923, 1.988, 2.000, 2.139, and 2.147 (6s, 6Ac), 4.773 (d, J 7.7 Hz, H-1⁴), 5.401 (s, PhCH); FAB-MS(M+Na)⁺ 1656. 4: $[\alpha]_D +105^\circ$ (c 1.9); δ_H 1.974, 1.980, 2.014, 2.020, 2.157, 2.162, 2.204 (7s, 6Ac and Lev), 5.122 (d, J 3.4 Hz, H-1²), 5.339 (dd, J 3.7 and 10.4 Hz, H-2¹), 5.538 and 5.419 (2d, J 2.4 and 1.9 Hz, H-4^{1,2}), 5.886 (d, J 9.8 Hz, NHAc), 6.585 (d, J 3.7 Hz, H-1¹), 8.683 (s, CNH). 6: $[\alpha]_D +97^\circ$ (c 0.4); δ_H 3.556 (dd, J 3.3 and 10.6 Hz, H-2), 4.036 (dd, J 2.2 and 10.3 Hz, Ser β H), 4.573 (m, Ser α H), 4.977 (d, J 2.6 Hz, H-1), 5.540 (s, PhCH). 9: M.p. 108-109°C; $[\alpha]_D +73^\circ$ (c 1.2); δ_H 2.040, 2.050, 2.134 (3s, 3Ac), 4.008 (dd, J 11.2 and 7.6 Hz, H-2¹), 4.500 and 4.185 (2d, J 11.9 Hz, CH₂CCl₃), 4.633 (d, J 7.6 Hz, H-1¹), 5.243 (d, J 3.4 Hz, H-1²), 5.476 (d, J 3.1 Hz, H-4²), 5.587 (s, PhCH). 10: M.p. 182-183°C; $[\alpha]_D +72^\circ$ (c 0.7); δ_H 2.007, 2.039, 2.132 and 2.196 (4s, 3Ac and Lev), 4.842 (d, J 8.1 Hz, H-1¹), 5.455 (dd, J 8.1 and 9.9 Hz, H-2¹), 5.488 (s, PhCH), 5.101 (d, J 4.0 Hz, H-1²), 5.741 (d, J 9.5 Hz, NHAc). 13: $[\alpha]_D -136.4^\circ$ (c 0.3); δ_H 2.123 (s, Lev), 5.558 (s, PhCH), 5.804 (dd, J 2.4 and 52.8 Hz, H-1). 14: $[\alpha]_D +133^\circ$ (c 0.3); δ_H 2.110 (s, Lev), 5.127 (dd, J 7.6 and 52.5 Hz, H-1), 5.535 (s, PhCH). 15: $[\alpha]_D +99^\circ$ (c 2.6); δ_H 0.148 and 0.184 (2s, 2CH₃), 0.926 (s, ^tBu), 5.543 (s, PhCH), 5.730 (dd, J 2.4 and 53.1 Hz, H-1). 16: $[\alpha]_D +23.5^\circ$ (c 0.5); δ_H 0.167, 0.130 (2s, 2CH₃), 0.926 (s, ^tBu), 4.022 (t, J 2.4 Hz, H-4), 5.058 (dd, J 7.6, 52.8 Hz, H-1), 5.541 (s, PhCH). 18: $[\alpha]_D +168^\circ$ (c 0.3); δ_H 2.116 (s, Lev), 5.033 (d, J 3.1 Hz, H-1), 5.506 (s, PhCH); δ_C 99.9 (C-1), 100.7 (PhCH). 19: $[\alpha]_D +72^\circ$ (c 1.5); δ_H 0.145 and 0.179 (2s, CH₃), 0.934 (s, ^tBu), 3.732 (dd, J 3.4 and 10.0 Hz, H-2), 4.470 (dd, J 7.3 and 10.7 Hz, H-3), 4.964 (d, J 3.4 Hz, H-1), 5.509 (s, PhCH). 20: $[\alpha]_D +29^\circ$ (c 0.7); δ_H 2.105 (s, Lev), 5.491 (s, PhCH); δ_C 102.3 (C-1), 100.9 (PhCH). 21: $[\alpha]_D +22^\circ$ (c 0.5); δ_H 0.117, 0.162 (2s, 2CH₃), 0.926 (s, ^tBu), 5.522 (s, PhCH). 22: $[\alpha]_D +101^\circ$ (c 0.3); δ_H 1.949, 1.959, 2.010, 2.034, 2.123, 2.149, 2.172 (7s, 6Ac and Lev), 4.699 (d, J 7.9 Hz, H-1²), 5.078 (d, J 3.7 Hz, H-1³), 5.209 (dd, J 7.6 and 10.1 Hz, H-2²), 5.514 (d, J 3.4 Hz, H-4²), 5.522 (s, PhCH); δ_C 96.3 (170.9 Hz, C-1³), 100.1 (172.1 Hz, C-1¹), 100.5 (162.3 Hz, PhCH), 102.1 (159.9 Hz, C-1²). 23: $[\alpha]_D +53^\circ$ (c 0.1); δ_H 1.991, 2.012, 2.018, 2.029, 2.159, 2.172 (6s, 6Ac), 4.982 (d, J 3.7 Hz, H-1¹), 5.029 (dd, J 3.0 and 11.3 Hz, H-3³), 5.040 (d, J 3.1 Hz, H-1³), 5.536 (s, PhCH). 25: $[\alpha]_D +42^\circ$ (c 0.1); δ_H 1.940, 1.972, 2.011, 2.022, 2.025, 2.076, 2.090, 2.113 (8s, 8Ac), 5.175 (d, J 3.7 Hz, H-1³), 5.440 (d, J 3.7 Hz, H-1⁴), 5.724 (d, J 9.5 Hz, SerNH or NHAc), 5.914 (d, J 8.6 Hz, SerNH or NHAc). FAB-MS (M+1)⁺ 1630. 26: $[\alpha]_D +82^\circ$ (c 0.1); δ_H 1.887, 1.901, 2.095, 2.033, 2.095, 2.121, 2.139 (7s, 7Ac), 2.158 (s, 2Ac), 5.192 (d, J 3.7 Hz, H-1³), 5.347 (d, J 3.1 Hz, H-1⁴). FAB-MS (M+1)⁺ 1648, (M+Na)⁺ 1670. 27: $[\alpha]_D +41.2^\circ$ (c 0.2, CH₃OH); $\delta_H(\text{CD}_3\text{OD})$ 1.889, 2.002 (2s, 2Ac). FAB-MS (M+1)⁺ 1090, (M+Na)⁺ 1112. 28: $[\alpha]_D +55.7^\circ$ (c 0.1, H₂O); $\delta_H(\text{D}_2\text{O at } 50^\circ)$ 1.222 (d, J 6.7 Hz, Fuc-CH₃), 2.068, 2.085 (2s, 2Ac), 2.710 (s, N(CH₃)₂), 4.713 (d, J 7.3 Hz, H-1²), 4.926 (d, J 3.7 Hz, H-1¹), 5.207 (d, J 4.0 Hz, H-1³), 5.295 (d, J 4.0 Hz, H-1⁴). FAB MS (M+1)⁺ 848, (M+Na)⁺ 870.
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